## Particulate organic matter and soil mineral nitrogen concentrations are good predictors of the soil nitrogen supply to canola following legume and non-legume crops in western Canada

M. St. Luce<sup>1,2</sup>, N. Ziadi<sup>2,10</sup>, B. J. Zebarth<sup>3</sup>, J. K. Whalen<sup>1</sup>, C. A. Grant<sup>4</sup>, E. G. Gregorich<sup>5</sup>, G. P. Lafond<sup>6†</sup>, R. E. Blackshaw<sup>7</sup>, E. N. Johnson<sup>8</sup>, J. T. O'Donovan<sup>9</sup>, and K. N. Harker<sup>9</sup>

 <sup>1</sup>Department of Natural Resource Sciences, Macdonald Campus, McGill University, 21, 111, Lakeshore Road, Ste-Anne-de-Bellevue, Quebec, Canada H9X 3V9; <sup>2</sup>Soils and Crops Research and Development Centre, Agriculture and Agri-Food Canada, 2560 Hochelaga Blvd., Quebec City, Quebec, Canada G1V 2J3;
<sup>3</sup>Potato Research Centre, Agriculture and Agri-Food Canada, 850 Lincoln Road, PO Box 20280, Fredericton, New Brunswick, Canada E3B 4Z7; <sup>4</sup>Brandon Research Centre, Agriculture and Agri-Food Canada, PO Box 1000A, R.R. #3, Brandon, Manitoba, Canada R7A 5Y3; <sup>5</sup>Eastern Cereal and Oilseed Research Centre, Agriculture and Agri-Food Canada, 960 Carling Ave, Ottawa, Ontario, Canada K1A 0C6; <sup>6</sup>Indian Head Research Farm, Agriculture and Agri-Food Canada, R.R. #1 Gov Road, PO Box 760, Indian Head, Saskatchewan, Canada S0G 2K0; <sup>7</sup>Lethbridge Research Centre, Agriculture and Agri-Food Canada, 5401 – 1 Avenue South, PO Box 3000, Lethbridge, Alberta, Canada T1J 4B1; <sup>8</sup>Saskatoon Research Centre, Agriculture and Agri-Food Canada, PO Box 10, Scott, Saskatoon, Saskatchewan, Canada S0K 4A0; and <sup>9</sup>Lacombe Research Centre, Agriculture and Agri-Food Canada, 6000 C and E Trail, Lacombe, Alberta, Canada T4L 1W1. Received 28 January 2013, accepted 3 September 2013. Published on the web 11 September 2013.

St. Luce, M., Ziadi, N., Zebarth, B. J., Whalen, J. K., Grant, C. A., Gregorich, E, G., Lafond, P., Blackshaw, R. E., Johnson, E. N., O'Donovan, J. T. and Harker, K. N. 2013. Particulate organic matter and soil mineral nitrogen concentrations are good predictors of the soil nitrogen supply to canola following legume and non-legume crops in western Canada. Can. J. Soil Sci. 93: 607–620. Accurate estimation of potential nitrogen (N) availability from preceding crops is essential to improve N fertilizer management in agricultural soils. Labile organic N fractions such as microbial biomass N (MBN), water-extractable organic N (WEON), particulate and light fraction organic matter N (POMN, LFOMN) are sensitive to management-induced changes and have the potential to predict N availability. This study assessed the impact of preceding legume [field pea (Pisum sativum L.), faba bean (Vicia faba L.), faba bean green manure] and non-legume crops [canola (Brassica napus L.) and wheat (Triticum aestivum L.)] on labile organic N fractions, mineral N (NH<sub>4</sub>-N+  $NO_3-N$ , potentially mineralizable N ( $N_0$ ) and soil N supply (canola grain yield and N uptake), and whether these soil parameters for the top 15 cm of soil could be used as indicators of soil N supply across no-till sites in western Canada. Labile organic N fractions and  $N_0$  were similar regardless of preceding crop. Soil N supply was greatest following faba bean green manure at four of five sites. POMN was the best single predictor of soil N supply ( $R^2 = 0.56$  and  $R^2 = 0.69$  for yield and N uptake, respectively). Soil N supply was primarily related to the combined effects of POMN, mineral N and sand content, which explained 68 and 71% of the variation in grain yield and N uptake, respectively. This study demonstrated that POMN and mineral N are relatively good predictors of soil N supply to canola in western Canada. Accounting for these parameters as well as soil texture may help improve N fertilizer recommendations for canola.

Key words: Canola, soil N tests, nitrogen supply, organic nitrogen, crop rotation

St. Luce, M., Ziadi, N., Zebarth, B. J., Whalen, J. K., Grant, C. A., Gregorich, E, G., Lafond, P., Blackshaw, R. E., Johnson, E. N., O'Donovan, J. T. et Harker, K. N. 2013. La matière organique particulaire et la concentration en azote minéral du sol sont de bons indicateurs de la teneur en azote disponible pour la culture du canola suivant un précédent cultural de cultures légumineuses ou non-légumineuses dans l'ouest canadien. Can. J. Soil Sci. 93: 607–620. L'estimation précise de l'azote (N) provenant d'un précédent cultural est essentielle à l'amélioration de la gestion de la fertilisation N dans les sols cultivés. Les fractions de N organique labile telles que le N de la biomasse microbienne (MBN), le N extractible à l'eau (WEON), la matière organique particulaire et la fraction légère (POMN, LFOMN) peuvent être utilisées pour prédire le N disponible du sol. Cette étude a évalué l'impact d'un précédent cultural de légumineuses [pois (*Pisum sativum* L.), féverole (*Vicia faba* L.), féverole comme engrais vert] et de non-légumineuses [canola (*Brassica napus* L.) et blé (*Triticum aestivum* L.)] sur les fractions de N organique labile, de N minéral (NH<sub>4</sub>-N+NO<sub>3</sub>-N), de N potentiellement minéralisable ( $N_0$ ) et N disponible du sol [rendement en grains et prélèvement de N par le canola] dans l'horizon 0–15 cm. Cette étude a aussi

<sup>†</sup>Deceased.

<sup>10</sup>Corresponding author (e-mail: noura.ziadi@agr.gc.ca).

Can. J. Soil Sci. (2013) 93: 607-620 doi:10.4141/CJSS2013-005

**Abbreviations:** LFOMN, light fraction organic matter N; MBN, microbial biomass N; N<sub>0</sub>, potentially mineralizable N; PCA, principal component analysis; POMN, particulate organic matter N; SON, soil organic N; WEON, water-extractable organic N

#### 608 CANADIAN JOURNAL OF SOIL SCIENCE

investigué si ces paramètres peuvent être utilisés comme indicateurs de N disponible du sol sous semis direct dans l'ouest canadien. Les fractions labiles de N et  $N_0$  ont été similaires quel que soit le précédent cultural. Le N disponible du sol a été plus élevé après un précédent de féverole dans quatre des cinq sites. Le POMN semble être le meilleur indicateur de N disponible du sol ( $R^2 = 0,56$  et  $R^2 = 0,69$  pour le rendement et le N exporté, respectivement). Le N disponible du sol a été principalement lié aux effets combinés du contenu en POMN, N minéral et la teneur du sol en sable, ce qui explique 68 et 71% de la variation du rendement en grain et de N exporté, respectivement. Cette étude a démontré que la teneur en POMN et N minéral sont relativement de bons indicateurs de N disponible du sol pour la culture du canola dans l'ouest canadien. Nous concluons que ces paramètres ainsi que la texture du sol peuvent aider à améliorer les recommandations d'engrais N pour la culture du canola.

Mots clés: Canola, analyse de sol, la fourniture d'azote, azote organique, rotation des cultures

The demand for canola grown in western Canada is increasing due to the healthy fatty acid balance in its edible oil and its suitability for biofuel and animal feed production (Canola Council of Canada 2012a). Given that canola is a major cash crop in western Canada that is mostly grown in rotations to reduce pest and disease build-up (Dosdall et al. 2012) and maintain soil fertility (Malhi et al. 2011a), increased yield per unit area is needed to meet this demand (Harker et al. 2012). Canola has a great N demand and therefore considerable N inputs are required (Malhi and Gill 2004; Cutforth et al. 2009). Previous findings suggest that inclusion of legumes in rotations can increase the soil N supply and possibly reduce N fertilizer inputs (Zentner et al. 2001; Soon and Arshad 2004; Lupwayi and Kennedy 2007; Campbell et al. 2011). Increased N availability following legumes is due to greater mineralizability of legume straw and roots, mostly due to their lower C:N ratio (Campbell et al. 1991b; Soon and Arshad 2004), and to the fact that legumes add N to soils through symbiotic fixation (Kumar and Goh 1999; Campbell et al. 2011). However, the amount of N from the legume crop that is available to a subsequent canola crop is unknown. The ability to accurately estimate this potential N availability would help improve N fertilizer recommendations and reduce external N inputs for canola production.

Prediction of soil N supply is challenging, since it varies temporally and spatially due to agricultural management history, soil properties and environmental conditions (St. Luce et al. 2011). A long-term aerobic incubation study proposed by Stanford and Smith (1972) to estimate potentially mineralizable N ( $N_0$ ) has long been recognized as the standard measure of the size of the mineralizable N pool. However, this method is not practical for routine use and may not represent actual N supply under field conditions (Malhi et al. 1992; Nyiraneza et al. 2012). Since the late 1960s, preplant soil NO<sub>3</sub>-N has been used as the basis for making N fertilizer recommendations in western Canada for both fallow and stubble crops (Soper and Huang 1963; Soper et al. 1971; Carson et al. 1974; Malhi et al. 1992) but it does not directly assess the important contribution of soil N supply originating from soil organic N (SON) (Walley et al. 2002).

Since a large proportion of SON is physically and chemically protected from microbial decomposition, it is the mineralizable (labile) SON that mainly contributes to soil N supply (Haynes 2005). Some SON fractions containing labile organic N include microbial biomass (MBN), water-extractable organic N (WEON), particulate and light fraction organic matter (POMN, LFOMN). Microbial biomass is primarily responsible for the decomposition of organic materials in soils and represents both a source (substrate) and sink (immobilization) of plant nutrients (Brookes 2001). Water-extractable organic N is frequently used as a substitute for soil solution collected in situ and includes the mobile and available portion of total dissolved organic N present in macropores and some smaller pores resulting from microbial decomposition of above- and below-ground residues (Chantigny 2003). Particulate organic matter and LFOM are composed of partially decomposed plant residues together with microbial by-products and are major sources of N for microbes (Gregorich et al. 2006). The LFOM, which contains appreciable quantities of carbohydrates and aliphatic compounds, is thought to be derived primarily from plant tissue whereas POM contains relatively more decomposed organic matter (Gregorich et al. 1996). Since these labile N fractions are actively involved in N cycling, it is hypothesized that they are an important component of soil N supply under field conditions. There are no reported studies relating these labile organic N fractions to soil N supply under field conditions following legume and non-legume crops in western Canada.

The objectives of this study were: (1) to assess the short-term impact of legume and non-legume preceding crops on soil MBN, WEON, POMN, LFOMN, mineral N,  $N_0$ , and field-based indices of soil N supply (canola yield and N uptake) and (2) to test the feasibility of using these parameters as predictors of soil N supply. This was evaluated using a series of five experimental sites across western Canada encompassing a range of soil properties under sub-humid and semi-arid conditions.

#### MATERIALS AND METHODS

#### Site Description

Samples used in this study were taken from an experiment established in 2009 at five sites across western Canada to investigate the influence of preceding legume and non-legume crops on soil fertility and canola production. The sites were at Brandon, Manitoba (lat. 50°02'N, long. 99°89'W); Indian Head, Saskatchewan (lat. 50°32'N, long. 103°40'W); Lacombe, Alberta (lat. 52°28'N, long. 113°44'W); Lethbridge, Alberta (lat. 49°41'N, long. 112°46'W); and Scott, Saskatchewan (lat. 52°21'N, long. 108°51'W). Soil characteristics and climatic conditions at each site are given in Tables 1 and 2, respectively. Before the experiment was initiated, the sites were cropped with either barley (*Hordeum vulgare* L.) or spring wheat.

## **Experimental Design**

The experiment was designed to accommodate a splitplot two-factor experiment in the subsequent year (2010) when canola was grown as the succeeding crop. Field pea, faba bean managed for grain, faba bean managed as green manure, canola and wheat were established as the main plots in 2009. Sowing rates of 50, 100, 150 and 300 seeds m<sup>2</sup> were used for faba bean, field pea, canola and wheat, respectively. The legumes were inoculated at each site. Fertilizer rates at each site are given in Table 3. Weeds were controlled with imazamox (20 g a.i. ha<sup>-</sup> with Merge<sup>®</sup> adjuvant added at 0.5% vol/vol. Following grain harvest, the straw of field peas, faba beans, canola and wheat were returned and evenly spread by hand over each plot and left on the soil surface. In comparison, the entire plant was returned to the soil (sprayed at the flat-pod stage with glyphosate (900 g a.e.  $ha^{-1}$ ) and clopyralid (50 g a.i.  $ha^{-1}$ ) and mowed in the fall if necessary) in the faba bean green manure plots. Straw yield and estimated crop residue N returned are provided in Table 3. No-till practices were used at all the sites.

In 2010, the main plots were split into five sub-plots as N rates. However, this study considered only the zero N rate. A small amount of N was applied in the control plots in the mono-ammonium phosphate that was used to supply P to canola at seeding (Table 3). The plot sizes differed according to location ( $2 \text{ m} \times 5 \text{ m}$  in Brandon,  $3.9 \text{ m} \times 10.7 \text{ m}$  in Indian Head,  $4.25 \text{ m} \times 12 \text{ m}$  in Lethbridge, and  $3.65 \text{ m} \times 15 \text{ m}$  in Lacombe and Scott), based on the seeding equipment available. Weeds were

burnt-off chemically prior to seeding canola. Hereafter, weeds were controlled with a post-emergence application of glyphosate at 450 g a.i.  $ha^{-1}$ .

### Plant Analysis

Canola grain yield and total N uptake from the zero N rate plots were used as field based measures of soil N supply. Subsamples of the harvested straw and grain were dried (25–30°C) and ground to pass a 1-mm screen. Grain yield was corrected to 10% moisture while straw yield was calculated on dry weight basis. Total N concentrations in straw and grain were determined by dry combustion with a Carlo-Erba C and N analyzer (Carlo-Erba Instruments, Milan, Italy). Total N uptake (kg N  $ha^{-1}$ ) was calculated as: (total N concentration in grain  $\times$  grain yield)+(total N concentration in straw  $\times$ straw yield). Total N uptake was subsequently corrected to field soil N supply by subtracting 75% of the starter N fertilizer input (Fox and Piekielek 1978; Hong et al. 1990) based on the assumption that 75% of the starter N was absorbed by the canola plant. Although this value was used for corn (Zea mays L.), there is no published value for canola.

## Soil Sampling

Soil samples from the 0- to 15-cm depth were a composite of two to four random samples per plot, collected from the control plots (0 kg N ha<sup>-1</sup>) in spring 2010 prior to the establishment of the canola crop. The samples were air-dried and passed through a 2-mm mesh sieve.

## Labile Organic N Fractions

Chloroform fumigation-direct extraction of incubated soil (Voroney et al. 2008) followed by persulphate digestion (Cabrera and Beare 1993) was used to determine the MBN concentrations. Briefly, 10 g of air-dried soil was incubated in a dark climate controlled chamber at 25°C and 60% water-filled pore space for 7 d. Subsequently, 5 g (dry weight) of soil was either directly extracted or fumigated for 24 h followed by extraction with 0.5 M K<sub>2</sub>SO<sub>4</sub> (1:4 soil:extractant). The persulphate reagent (50 g K<sub>2</sub>S<sub>2</sub>O<sub>8</sub>+30 g H<sub>3</sub>BO<sub>4</sub>+15 g NaOH in 1 L deionized water) was added to the 0.5 M K<sub>2</sub>SO<sub>4</sub> extracts

Table 1. Soil characteristics at five sites in 2009 before the establishment of legume and non-legume preceding crops									
Site	Preceding Crop	$\text{Total } N^{\textbf{z}}$	$\text{Organic } C^{z}$	Sand <sup>y</sup>	Silt <sup>y</sup>	Clay <sup>y</sup>	Textural class	Soil type	$\mathrm{pH}^{\mathbf{x}}$
(g kg <sup>-1</sup> )									
Brandon	Barley	3.1	31	486	255	259	Sandy Clay Loam	Black Chernozem	7.5
Indian Head	Wheat	2.5	26	64	275	661	Heavy Clay	Orthic Black Chernozem	7.3
Lacombe	Barley	4.7	47	395	340	265	Loam	Black Chernozem	6.4
Lethbridge	Wheat	1.9	14	308	462	230	Loam	Dark Brown Chernozem	7.8
Scott	Wheat	2.2	23	404	401	195	Loam	Dark Brown Chernozem	5.7

<sup>z</sup>Dry combustion using an Elementar CN Analyzer (Elementar Analysensysteme GmbH, Hanua, Germany). Carbonates were removed before analysis (Skjemstad and Baldock 2008).

<sup>y</sup>Soil texture was determined using the pipette method after organic matter removal (Kroetsch and Wang 2008).

<sup>x</sup>1:1 soil: water slurry (Hendershot et al. 2008).

Table 2. Total precipitation and mean air temperature during the growing season (May – September) in 2009 and 2010 at the five sites in comparison to the 30-y average  $(1971-2000)^{z}$ 

	Total precipitation (mm)			Mean air temperature (°			
Site	2009	2010	30-yr avg.	2009	2010	30-yr avg.	
Brandon Indian	298 233	446 293	322 296	14.6	14.6	15.0	
Head	232	506	320	13.1	11.8	12.9	
Lethbridge Scott	241 218	394 497	249 244	15.5 13.6	14.1 13.0	15.0 14.0	

<sup>&</sup>lt;sup>z</sup>Environment Canada (climate.weatheroffice.gc.ca/climate\_normals/ results\_e.html).

(ratio 1:1) in a Kimax glass tube, capped and autoclaved at 120°C for 30 min. The MBN was subsequently calculated as [(total N in digests of fumigated soil extracts – total N in non-fumigated soil extracts)/ $k_{\rm EN}$ ] where  $k_{\rm EN}$ is the extraction coefficient 0.54 (Brookes et al. 1985).

For WEON extraction, 15 mL of 5 mM calcium chloride (CaCl<sub>2</sub>) was added to 7.5 g of air-dried soil (1:2 soil:solution ratio) and gently stirred at room temperature for 1 min followed by centrifugation at 9000 rpm for 10 min (Chantigny et al. 2008). The supernatant was

then filtered through a vacuum filter unit equipped with a 0.45  $\mu$ m nylon membrane filter (Pall Corp., Ann Arbor, MI). The WEON concentration was subsequently calculated as the difference between the total dissolved N and the mineral N concentration (NH<sub>4</sub>-N+ NO<sub>3</sub>-N), which were quantified on a QuickChem 8000 Lachat autoanalyzer (Lachat Instruments, Loveland, CO).

A modification of the procedure described by Gregorich and Beare (2008) was used to separate the POM. Briefly, 25 g of air-dried soil was dispersed by shaking in 100 mL of deionized water and 10 glass beads for 16 h. The dispersed soil was passed through a 53-µm sieve. The retained sand and macroorganic matter were dried at 50°C, weighed and then ground using a mortar and pestle to pass through a 250-µm sieve. The density fractionation procedure was used to separate the LFOM (Gregorich and Beare 2008) from 25 g air-dried soil by shaking in a sodium iodide solution (specific gravity of 1.8 g cm<sup>-3</sup>) for 1 h and allowing the soil mineral particles to settle for 48 h before recovering the suspended LFOM. The N contents of the POM and LFOM were determined by dry combustion using an Elementar CN Analyzer (Elementar Analysensysteme GmbH, Hanua, Germany).

Crop	Brandon	Indian Head	Lacombe	Lethbridge	Scott						
	Fertilizer rates (kg ha <sup>-1</sup> ) at seeding in 2009										
Legumes	18 kg N	6 kg N	6 kg N	6 kg N	4 kg N						
0	$25 \text{ kg } P_2O_5$	$26 \text{ kg } P_2O_5$	$26 \text{ kg } P_2O_5$	$26 \text{ kg } P_2O_5$	18 kg P <sub>2</sub> O <sub>5</sub>						
	15 kg S	0	0	0	0 - 1						
Canola	69 kg N	81 kg N	118 kg N	82 kg N	102 kg N						
	$20 \text{ kg } P_2O_5$	$26 \text{ kg } P_2O_5$	$38 \text{ kg } P_2O_5$	$23 \text{ kg } P_2O_5$	16 kg P <sub>2</sub> O <sub>5</sub>						
	15 kg S		76 kg K <sub>2</sub> O		12 kg K <sub>2</sub> O						
	-		10 kg S		7 kg S						
Wheat	70 kg N	44 kg N	95 kg N	44 kg N	82 kg N						
	$30 \text{ kg } P_2 O_5$	$26 \text{ kg } P_2O_5$	$26 \text{ kg } P_2O_5$	15 kg S	18 kg P <sub>2</sub> O <sub>5</sub>						
	15 kg S		52 kg $K_2O$	-	15 kg S						
	-		7 kg S		-						
		Sti	raw yield in 2009 (kg ha <sup>-</sup>	-1)							
Canola	3700	6688	9088	6091	5348						
Faba Bean	2215	3617	6765	2139	4615						
Field Pea	4462	5237	5035	4095	$ND^{z}$						
Green Manure	3262	4596	5033	5247	4296						
Wheat	3264	6349	7152	1278	6348						
		Estimated crop	residue N returned in 200	9 $(kg \ N \ ha^{-1})$							
Canola	27	19	45	22	29						
Faba Bean	29	27	60	16	29						
Field Pea	51	54	72	29	ND						
Green Manure	64	140	141	157	115						
Wheat	26	30	27	11	30						
		Fertilizer	rates (kg ha <sup>-1</sup> ) at seeding	ig in 2010							
Canola	15 kg N	6 kg N	7 kg N	7 kg N	6 kg N						
	30 kg P <sub>2</sub> O <sub>5</sub>	$29 \text{ kg } P_2O_5$	31 kg P <sub>2</sub> O <sub>5</sub>	13 kg P <sub>2</sub> O <sub>5</sub>	29 kg P <sub>2</sub> O <sub>5</sub>						
	10 kg S	$47 \text{ kg } \text{K}_2\text{O}$	31 kg K <sub>2</sub> O	26 kg K <sub>2</sub> O	33 kg K <sub>2</sub> O						
	-	16 kg S	10 kg S	10 kg S	10 kg S						

<sup>z</sup>Not determined.

#### Potentially Mineralizable N

A 24-wk aerobic incubation procedure, based on the method of Curtin and Campbell (2008), was used to estimate  $N_0$ . Briefly, soils were mixed with an equal amount of sand for light-textured soils and twice the amount of sand for fine-textured soils, packed into leaching tubes, and wetted to 55% of the soil's waterholding capacity by leaching with 100 mL of 0.01 M CaCl<sub>2</sub> followed by suction ( $\sim 10$  kPa) and incubated at 25°C for 24 wk. Thereafter, the soil moisture content was kept at 55% of the water-holding capacity. The first leaching represented the initial NO<sub>3</sub>-N and NH<sub>4</sub>-N concentrations. The soils were leached every 2 wk for the first 12 wk and every 4 wk thereafter with 100 mL 0.01 M CaCl<sub>2</sub> followed by a 25-mL zero-N nutrient solution (Curtin and Campbell 2008). Leachates were filtered through a Whatman No. 42 filter paper and analyzed colorimetrically for NO<sub>3</sub>-N and NH<sub>4</sub>-N (Zebarth and Milburn 2003). Mineral N in the sum of  $NO_3$ -N+NH<sub>4</sub>-N in the first 2-wk period, herein considered as Pool I (Sharifi et al. 2007b), was not used in the curve fitting procedure because it represented the initial flush of mineralization upon rewetting. Cumulative N mineralized between weeks 2 and 24 represented total mineralized N and was referred to as Pool II (Sharifi et al. 2007b).

To estimate  $N_0$ , the following first order kinetic model was fitted to the data using the Marquardt iteration method:

$$N_{\min} = N_0 [1 - \exp(-kt)]$$

where  $N_{min}$  is the cumulative amount of N mineralized (mg N kg<sup>-1</sup>) at time *t*,  $N_0$  is the potentially mineralizable N (mg N kg<sup>-1</sup>) and *k* is mineralization rate coefficient (wk<sup>-1</sup>). The model was fitted to data using both fixed and unfixed *k* approaches (Wang et al. 2003). The fixed *k* approach (k = 0.054 wk<sup>-1</sup>) substantially reduced the coefficient of variation of the  $N_0$  data from 94 to 40% and was therefore used in this study. All measurements were expressed as concentration per kilogram of soil.

#### Statistical Analyses

All statistical analyses were performed using SAS software (SAS Institute, Inc. 2004). Data were checked for normality based on the Shapiro–Wilk test and transformed when appropriate. Analysis of variance was performed using the GLM procedure based on a randomized complete block design to evaluate the effect of each preceding crop on soil MBN, WEON, POMN, LFOMN, mineral N,  $N_0$ , mineralizable N pools, and canola grain yield and N uptake in control plots (N uptake minus starter N fertilizer applied in 2010) for each site. When significant (P < 0.05) effects were observed, means were compared with a post-hoc Least Significant Difference (LSD) test at P < 0.05. It was hypothesized that legumes would increase the concentration of labile organic N fractions and contribute more to soil N supply than non-legumes; therefore, a pre-planned contrast was constructed to distinguish legume and non-legume effects.

Single linear regression using PROC REG was used to assess the proportion of the variability in yield and N uptake that could be explained by each of the labile organic N fractions, mineral N,  $N_0$ , Pool I, Pool II and other soil chemical and physical properties. The linear model was used since it provided a better fit than the quadratic model. Multivariate analysis was also used to explain the variability in soil N supply. Since climatic conditions play a significant role in N dynamics and crop performance, mean air temperature and total precipitation for the 2010 growing season were included in the analysis. The data were subjected to the Kaiser's measure of sampling adequacy (MSA) to detect unique correlations (Kaiser 1974). Small values of MSA indicate that the correlation between paired variables are unique, that is, not related to remaining variables outside of each simple correlation (Wuensch 2004). Kaiser (1974) described MSA's < 0.6 not suitable for principal component analysis (PCA). The MSA of the entire data set was 0.5 and therefore was not suitable for the PCA. Because clay, sand and silt, as well as mineral N (NO<sub>3</sub>-N+NH<sub>4</sub>-N), NO<sub>3</sub>-N and NH<sub>4</sub>-N were highly intercorrelated, silt, NO<sub>3</sub>-N and NH<sub>4</sub>-N were removed from the data set. The recalculated MSA was 0.67; hence the PCA could be performed. The PCA, using the PROC FACTOR of SAS, was then used to develop artificial variables (PCs) that accounted for much of the variance contained in the variables. The retained PCs (eigenvalues >1) were rotated orthogonally with the VARIMAX option of SAS (Nyiraneza et al. 2009). A soil parameter was assigned to a PC for which it had the highest loading value. The scoring coefficients were obtained using the SCORE procedure of SAS software. Stepwise regression was then conducted using canola yield or N uptake as the dependent variables and the scoring coefficients of the retained PCs as the independent variables. The significance level to enter the model was P = 0.15 and the significance to stay was  $P \le 0.05$ .

## RESULTS

#### Soil Characteristics and Climatic Conditions

Surface soils at the experimental sites varied in soil texture and were classified as loam at Lacombe, Lethbridge and Scott, sandy clay loam at Brandon and heavy clay at Indian Head (Table 1). Soil total N and organic C ranged from 1.9 to 4.7 g N kg<sup>-1</sup> and 26 to 47 g C kg<sup>-1</sup>, respectively. The soils tended to be slightly acidic to calcareous with pH >7.0 at three of the five sites. Precipitation during the growing season in 2009, during growth of the preceding crops, ranged from 73 to 97% of climate normals (Table 2). In comparison, in 2010, growing season precipitation was near normal at Indian Head but was above normal at other sites including more than 50% above normal at Lacombe and Lethbridge

Site	Preceding crop	MBN	WEON	POMN	LFOMN	MinN	$N_0$	Pool I	Pool II	
		$(m\sigma N k\sigma^{-1})$								
Brandon	Canola	45 (2) <sup>y</sup>	5.8 (0.7)	173 (16)	50 (6)	10.3 (0.9)	59bc (4)	29 (2)	32 (3)	
	Faba bean	47 (3)	5.8 (0.4)	181 (17)	56 (3)	10.6 (1.8)	70ab (3)	19 (2)	37 (3)	
	Field pea	39 (2)	5.3 (0.2)	176 (7)	45 (5)	12.2 (1.2)	55c (3)	28 (2)	30 (2)	
	Green manure	42 (3)	6.0 (0.5)	181 (21)	44 (3)	11.1 (0.7)	76a (7)	21 (6)	41 (4)	
	Wheat	47 (2)	5.3 (0.4)	197 (22)	55 (6)	10.1 (1.4)	60bc (4)	30 (5)	34 (3)	
	<i>P</i> value	NS	NS	NS	NS	NS	*	NS	NS	
	Contrast <sup>*</sup>	NS	NS	NS	NS	NS	NS	NS	NS	
Indian Head	Canola	51 (1)	3.9 (0.2)	214 (13)	91 (28)	5.1c (0.3)	63 (10)	8 (2)	32 (2)	
	Faba bean	47 (3)	4.0 (0.1)	203 (13)	63 (25)	6.1bc (0.2)	51 (1)	11 (3)	30 (0.3)	
	Field pea	43 (3)	4.3 (0.2)	178 (22)	52 (7)	9.3a (1.7)	79 (6)	23 (2)	46 (2)	
	Green manure	48 (2)	4.4 (0.3)	174 (38)	64 (18)	9.9a (0.7)	66 (4)	14 (2)	36 (4)	
	Wheat	49 (2)	4.2 (0.3)	229 (20)	79 (20)	8.4ab (1.1)	60 (11)	16 (6)	38 (8)	
	<i>P</i> value	NS	NS	NS	NS	*	NS	NS	NS	
	Contrast	NS	NS	*	NS	NS	NS	NS	NS	
Lacombe	Canola	57 (2)	4.2 (0.7)	634 (76)	48 (7)	11.0 (3.5)	97 (10)	30 (6)	48 (4)	
	Faba bean	41 (10)	4.9 (1.0)	600 (50)	53 (15)	10.7 (0.5)	109 (19)	22 (9)	57 (9)	
	Field pea	47 (8)	4.5 (1.2)	610 (69)	49 (7)	11.5 (3.4)	115 (18)	34 (11)	58 (6)	
	Green manure	32 (6)	5.7 (0.6)	643 (100)	77 (15)	16.4 (1.4)	139 (21)	53 (11)	80 (11)	
	Wheat	41 (4)	5.5 (0.9)	618 (69)	51 (10)	8.8 (1.4)	112 (12)	39 (4)	63 (7)	
	<i>P</i> value	NS	NS	NS	NS	NS	NS	NS	NS	
	Contrast	NS	NS	NS	NS	*	NS	NS	NS	
Lethbridge	Canola	54 (4)	5.0 (0.8)	212 (22)	41 (9)	6.8 (0.6)	79 (24)	12 (2)	45 (15)	
	Faba bean	51 (3)	4.0 (0.3)	187 (17)	25 (5)	8.4 (1.2)	80 (22)	17 (2)	52 (14)	
	Field pea	57 (5)	5.1 (1.2)	290 (62)	47 (11)	8.7 (0.8)	85 (23)	11 (4)	48 (13)	
	Green manure	47 (1)	4.6 (0.3)	268 (53)	42 (14)	11.8 (1.9)	80 (21)	18 (2)	51 (15)	
	Wheat	48 (4)	4.1 (0.3)	237 (43)	49 (7)	8.4 (1.1)	90 (9)	11 (4)	44 (8)	
	<i>P</i> value	NS	NS	NS	NS	NS	NS	NS	NS	
	Contrast	NS	NS	NS	NS	NS	NS	NS	NS	
Scott	Canola	30 (1)	5.9 (1.1)	295 (51)	126 (18)	6.7b (0.8)	132 (14)	30 (4)	68 (10)	
	Faba bean	25 (2)	5.9 (0.3)	283 (21)	136 (12)	6.5b (0.7)	127 (11)	34 (5)	68 (8)	
	Field pea	28 (2)	5.4 (0.5)	235 (38)	104 (19)	7.7b (1.3)	106 (12)	29 (6)	60 (6)	
	Green manure	29 (3)	6.4 (0.4)	286 (12)	123 (7)	13.0a (0.7)	123 (10)	38 (1)	69 (7)	
	Wheat	32 (3)	5.6 (0.6)	382 (31)	145 (5)	8.4b (0.2)	133 (10)	33 (1)	77 (6)	
	<i>P</i> value	NS	NS	NS	NS	***	NS	NS	NS	
	Contrast	NS	NS	*	NS	NS	NS	NS	NS	

Table 4. Concentration of labile organic nitrogen fractions, mineral N and mineralizable N pools<sup>z</sup> at the 0- to 15-cm depth in spring 2010. The preceding crops were harvested in fall 2009 and crop residues left on the soil surface in a no-till system

<sup>2</sup>MBN, microbial biomass N; WEON, water-extractable organic N; POMN, particulate organic matter N; LFOMN; light fraction organic matter N; MinN, initial mineral N ( $NH_4$ -N+ $NO_3$ -N);  $N_o$ , potentially mineralizable N; Pool I, cumulative amount of N mineralized in the first 2 wk following rewetting; Pool II, cumulative amount of N mineralized between 2 and 24 wk. <sup>9</sup>Standard error of mean (n =4).

<sup>x</sup>Legumes vs. non-legumes.

a-c Means followed by different lower case letters are significantly different at P < 0.05.

\*, \*\* Significant at P < 0.05 and P < 0.001, respectively; NS, not significant at P < 0.05.

and over 100% above normal at Scott. Mean air temperature during the growing season was within 0.5°C of the climate normal in all cases except for Scott, where the mean air temperature was 1.4°C below normal in both years. At each site, the straw yield was generally greater for the preceding non-legumes (canola and wheat), while the estimated crop residue N was greater for preceding faba bean green manure than the other preceding crops (Table 3).

#### Labile Organic N Fractions

The POMN was the largest fraction measured in this study, accounting for 5.7 to 12.7% of total soil N (Table 4). The MBN and LFOMN accounted for 0.9 to 2.7% and 1.1 to 5.3% of total N, respectively, whereas WEON

accounted for about 0.2%. There was no effect (P > 0.05) of preceding crop on soil MBN, WEON, POMN or LFOMN at any site (Table 4). Contrast analysis showed that POMN was significantly (P < 0.05) greater following preceding non-legume than legume crops at Indian Head and Scott.

## Soil Mineral N

Soil mineral N concentration ranged from 5.1 to 16.4 mg N kg<sup>-1</sup> in the 0- to 15-cm layer (Table 4). Soil NO<sub>3</sub>-N was the dominant form of mineral N at these sites (data not shown), accounting for more than 80% of the total mineral N at Brandon, Indian Head and Lethbridge, and between 60 and 70% at Lacombe and Scott. Significant differences in soil mineral N concentration

among the preceding crops were observed at only two (Indian Head and Scott) of the five sites (Table 4). At Indian Head, soil mineral N was greater for preceding faba bean green manure and field pea crops than for preceding faba bean grain and canola crops (Table 4). At Scott, soil mineral N was greater for a preceding faba bean green manure than for all other preceding crops. Contrast analysis showed significantly greater soil mineral N concentrations following preceding legume than non-legume crops at Lacombe.

#### Mineralizable N Pools

 $N_0$  ranged from 51 to 139 mg N kg<sup>-1</sup> and represented 2 to 5% of total soil N. Pools I and II represented an average of 0.8 and 1.8% of total soil N, respectively. The  $N_0$  varied significantly among the preceding crops only at Brandon (Table 4). At this site,  $N_0$  was greater for a preceding faba bean green manure crop than for preceding canola, field pea and wheat crops (Table 4).

#### Soil N Supply

Canola grain yield from unfertilized plots ranged from 582 to 4283 kg ha<sup>-1</sup>, while N uptake in the above-ground plant ranged from 26 to 229 kg N ha<sup>-1</sup>. Canola grain yield varied significantly among preceding crops at all sites except Brandon (Table 5). At Indian Head, canola grain yield was greater for preceding faba bean green manure crop than faba bean grain, canola and wheat crops. At Lacombe, canola grain yield was greater for preceding faba bean green manure crop than field pea, canola and wheat crops. Canola grain yield at Lethbridge was greater for preceding faba bean green manure than canola, while at Scott, canola grain yield was greater for a preceding faba bean green manure than all other crops. A comparison between preceding legume and nonlegume crops showed that canola grain yield was greater following legumes than non-legumes at Lacombe and Lethbridge.

Crop N uptake varied significantly among the preceding crops at three (Indian Head, Lacombe and Scott) sites (Table 5). At Indian Head, canola N uptake was greater for preceding faba bean green manure crop than faba bean grain, canola and wheat crops. At Lacombe, canola N uptake was greater for preceding faba bean green manure crop than field pea and canola crops. Canola grain yield at Scott was greater for a preceding faba bean green manure than all other preceding crops. Contrast analysis showed significantly greater canola N uptake following preceding legume than non-legume crops at Scott.

# Relationship Between Soil N Supply and Individual Soil Parameters

Canola grain yield was related to N uptake (r = 0.97, P < 0.001, data not shown). The best single predictor of soil N supply was POMN ( $R^2 = 0.56$  and  $R^2 = 0.69$  for grain yield and crop N uptake, respectively; Fig. 1). Soil mineral N concentration (Fig. 1) and total soil N

Table 5. Canola grain yield and N uptake in 2010 following legume and non-legume crops at five sites. The preceding crops were harvested in fall 2009 and crop residues left on the soil surface in a no-till system

Site	Preceding crop	Grain yield (kg ha <sup>-1</sup> )	N uptake (kg N ha <sup>-1</sup> )
Brandon	Canola Faba bean Field pea Green manure Wheat <i>P</i> value Contrast <sup>y</sup>	1505 (221) <sup>z</sup> 1719 (217) 1852 (243) 2000 (324) 1525 (410) NS NS	56 (13) 67 (11) 71 (12) 73 (16) 64 (17) NS NS
Indian Head	Canola Faba bean Field pea Green manure Wheat <i>P</i> value Contrast	1041 (416) <i>bc</i> 582 (56) <i>c</i> 1244 (38) <i>ab</i> 1621 (90) <i>a</i> 879 (150) <i>bc</i> ** NS	42 (15)bc 26 (6)d 54 (4)ab 74 (3)a 43 (11)bc * NS
Lacombe	Canola Faba bean Field pea Green manure Wheat <i>P</i> value Contrast	2635 (334)b 3457 (123)ab 2766 (444)b 4283 (52)a 2772 (657)b *	125 (8) <i>c</i> 199 (15) <i>ab</i> 141 (19) <i>bc</i> 229 (26) <i>a</i> 161 (39) <i>ab</i> * NS
Lethbridge	Canola Faba bean Field pea Green manure Wheat <i>P</i> value Contrast	1547 (86)c 1990 (49)abc 2383 (323)ab 2637 (312)a 1979 (176)abc *	64 (7) 80 (4) 114 (24) 117 (22) 90 (11) NS NS
Scott	Canola Faba bean Field pea Green manure Wheat <i>P</i> value Contrast	1253 (194)b 1134 (259)b 1398 (172)b 2100 (124)a 1185 (143)b * NS	50 (6)b 65 (14)b 61 (3)b 97 (7)a 58 (11)b *

<sup>z</sup>Standard error of mean (n = 4).

<sup>y</sup>Legumes vs. non-legumes.

a-c Means followed by different lower case letters are significantly different at P < 0.05.

\*, \*\* Significant at P < 0.05 and P < 0.01, respectively; NS, not significant at P < 0.05.

(Table 6) accounted for similar variations in N uptake but mineral N gave a slightly better prediction for yield. The  $N_0$  and Pool II were poor predictors of soil N supply, explaining <25 and <30% of the variability in yield and N uptake, respectively (Table 6). Pool I explained only 22 and 25%, respectively, of the variation in canola grain yield and N uptake.

## Prediction of Soil N Supply Using Multivariate Analysis

The PCA retained four PCs, accounting for 92% of the total variance (Table 7). The retained PCs explained >80% of all parameters, except mineral N. The first PC accounted for 49% of the total variance and had positive loadings for LFOMN, WEON and  $N_0$ , and negative



Fig. 1. Relationships between canola grain yield and N uptake and particulate organic matter N (POMN) (a and b) and initial mineral N (c and d) at the 0- to 15-cm depth. Initial mineral N ( $NH_4-N+NO_3-N$ ) extracted with 0.01 M CaCl<sub>2</sub> prior to the start of the long-term incubation.

loadings for MBN and soil pH (Table 7). About 18% of the total variance was explained by PC<sub>2</sub> and was mainly composed of POMN, total soil N and mean air temperature. The third PC explained 15% of the total variance and had positive loading values for sand and total precipitation, and a negative value for clay. The fourth PC explained 10% of the total variance and was mainly composed of mineral N, Pool I and C:N ratio. Stepwise regression analysis was used to relate canola grain yield and N uptake, respectively, as the dependent variables and the retained PCs as independent variables. The combined PCs explained 75 and 71%, respectively, of the variations in canola grain yield and N uptake (Table 8). While three of the four PCs were significant to remain in the model for N uptake, all four PCs remained in the model for grain yield (Table 8). The  $PC_2$  was the first selected in each model, accounting for 30 and 44%, respectively of canola grain yield and N uptake (Table 8). This was followed by PCs 3, 4 and 1 for grain yield and PCs 3 and 4 for N uptake. The proportion of the total variability of grain yield and N uptake explained by PC<sub>3</sub> was 20 and 15%, respectively, while the proportion explained by PC<sub>4</sub> was 18 and 12%, respectively.

## DISCUSSION

## Canola Grain Yield and N Uptake

The trend towards greater canola grain yield at four of five sites and greater N uptake at three of five sites following preceding faba bean green manure versus other preceding legume and non-legume crops suggests that faba bean green manure can reduce the need for external N inputs in canola production. This result is

	Cano	la grain yield (kg h	$a^{-1}$ )	Canola N uptake (kg N ha <sup><math>-1</math></sup> )			
Independent variable <sup>z</sup>	Intercept	slope	$R^2$	Intercept	slope	$R^2$	
LFOMN (mg N kg $^{-1}$ )	2485	-8.53	0.11	112	-0.34	0.05	
MBN (mg N kg <sup><math>-1</math></sup> )	1860	0.92	0.00	102	-0.32	0.00	
WEON (mg N kg $^{-1}$ )	1130	153	0.02	51	7.54	0.01	
$N_0 \text{ (mg N kg}^{-1})$	673	13.7	0.20*	4.59	0.94	0.27**	
Pool I (mg N kg <sup><math>-1</math></sup> )	1018	36.1	0.22**	33.2	2.28	0.25*	
Pool II (mg N kg $^{-1}$ )	697	24.2	0.18*	5.75	1.67	0.25*	
Clay (mg kg <sup><math>-1</math></sup> )	3036	-4.66	0.28**	148	-0.24	0.22*	
Silt $(mg kg^{-1})$	966	2.19	0.02	46	0.10	0.01	
Sand (mg kg <sup><math>-1</math></sup> )	892	3.04	0.15*	36	0.16	0.13	
Total N (g N kg <sup><math>-1</math></sup> )	386	502	0.40***	-8.41	32.3	0.47***	
рН	2690	-113	0.02	180	-13.05	0.06	

Table 6. Linear regressions relating labile organic N fractions, N<sub>0</sub>, mineralizable N pools and other soil properties at the 0- to 15-cm depth to canola grain yield and N uptake following legume and non-legume crops across five sites

<sup>2</sup>LFOMN, light fraction organic matter N; MBN, microbial biomass N; WEON, water-extractable organic N; N<sub>0</sub>, potentially mineralizable N; Pool I, cumulative amount of N mineralized in the first 2 wk following rewetting; Pool II, cumulative amount of N mineralized between 2 and 24 wk. \*, \*\*, \*\*\* Significant at P < 0.05, P < 0.01, and P < 0.001, respectively.

partly due to the larger amount of crop residue N returned by the preceding faba bean green manure crop compared with the other crops where the seeds were harvested. Although straw yield of the faba bean green manure crop was lower than canola, wheat and field pea at all sites, except that field pea was greater at Lethbridge, only the straw of field pea, canola and wheat were returned as crop residues, while the entire faba bean plant was present as crop residues in the faba bean green manure plots. The presence of a greater amount of N-rich residues may have caused higher net N mineralization in the faba bean green manure plots during the growing season compared with the other

Table 7. Rotated principle components (PCs) and total communality estimates (CE) for prediction of soil N supply to canola following legume and non-legume crops across five sites

Parameters <sup>z</sup>	$PC_1$	$PC_2$	$PC_3$	$PC_4$	CE
$\overline{\text{LFOMN} (\text{mg N kg}^{-1})}$	0.94				0.92
POMN (mg N kg <sup><math>-1</math></sup> )		0.93			0.98
MBN (mg N kg <sup><math>-1</math></sup> )	-0.91				0.90
WEON (mg N kg $^{-1}$ )	0.59				0.88
MinN (mg N kg <sup><math>-1</math></sup> )				0.78	0.70
$N_0 (\text{mg N kg}^{-1})$	0.61				0.92
Pool I (mg N kg <sup><math>-1</math></sup> )				0.62	0.89
Clay $(g kg^{-1})$			-0.90		0.95
Sand $(g kg^{-1})$			0.97		0.97
Total N (g N kg <sup><math>-1</math></sup> )		0.78			0.94
pH	-0.86				0.99
Mean Temperature (°C)		-0.93			0.97
Total Precipitation (mm)			0.72		0.94
C/N				0.58	0.85
Eiegenvalues	6.87	2.53	2.05	1.36	
Variance proportion	0.49	0.18	0.15	0.10	
Cumulative variance	0.49	0.67	0.82	0.92	

<sup>z</sup>LFOMN, light fraction organic matter N; POMN, particulate organic matter N, MBN, microbial biomass N; WEON, water-extractable organic N; MinN, initial mineral N (NH<sub>4</sub>-N+NO<sub>3</sub>-N);  $N_0$ , potentially mineralizable N; Pool I, cumulative amount of N mineralized in the first 2 wk following rewetting; C/N, soil C/N ratio.

plots and mineralization may have varied according to soil properties and climatic conditions across the sites. No information on the quantity and quality of belowground plant residues was collected in this study but Unkovich and Pate (2000) reported that about 40% of the total plant N in faba bean is found below ground. The greater  $N_2$ -fixation potential of faba bean (12–330 kg  $N_2$  ha<sup>-1</sup> yr<sup>-1</sup>) (Lupwayi and Kennedy 2007) than field pea may also account for these results. It was estimated that field pea fixed about 90 kg  $N_2$  ha<sup>-1</sup> yr<sup>-1</sup> in the Canadian prairies (Biederbeck et al. 1996), while in Australia, Unkovich and Pate (2000) reported that faba bean fixed 144 kg  $N_2$  ha<sup>-1</sup> in comparison with 96 kg  $N_2$  ha<sup>-1</sup> by field pea. It should also be noted that a considerable amount of N is removed from the soilplant system in legume crops when they are harvested for seed, unlike when grown as green manure crop. Many studies reported increased N availability following legumes (Soon et al. 2001; Campbell et al. 2011; Zotarelli et al. 2012) as a result of faster decomposition of N-rich legume residues. In our study, canola grain yield was greater following legumes than non-legumes at Lacombe and Lethbridge only, while N uptake was

Table 8. Stepwise regression models relating canola grain yield and N uptake to retained PCs  $% \left( {{{\rm{T}}_{{\rm{S}}}} \right)$ 

Equation	$R^{2\mathbf{z}}$
Canola grain yield $(kg ha^{-1})$	
$Y = 1899 + 460 \times PC_2$	0.30
$Y = 1899 + 460 \times PC_2 + 384 \times PC_3$	0.50
$Y = 1899 + 460 \times PC_2 + 384 \times PC_3 + 358 \times PC_4$	0.68
$Y = 1899 + 460 \times PC_2 + 384 \times PC_3 + 358 \times PC_4 - 225 \times PC_1$	0.75
Canola N uptake (kg N ha <sup>-1</sup> )	
$Y = 89 + 33 \times PC_2$	0.44
$Y = 89 + 33 \times PC_2 + 19 \times PC_3$	0.59
$Y = 89 + 33 \times PC_2 + 19 \times PC_3 + 18 \times PC_4$	0.71

<sup>z</sup>All models were significant at P < 0.001

greater following legumes at Scott only. This suggests that the relative benefit of legumes in crop rotations may not be obvious in the short-term and depends on the specific legume species, the quality of the legume residue, the legume biomass yield, whether the legume is grown for seed or as a green manure crop since considerable N is removed in the harvested seed, and the effect of soil and climatic conditions (Kumar and Goh 1999; Thorup-Kristensen et al. 2003).

### Labile Organic N Fractions and Mineral N

A major goal of this study was to determine how residues from the preceding crops affected other components of the soil N supply, namely soil MBN, POMN, LFOMN, WEON and soil mineral N concentrations. This study showed no influence of preceding crops on soil MBN, WEON, POMN and LFOMN at any sites. These labile organic N fractions are known to be relatively variable and responsive to changes in management (Griffin and Porter 2004; Haynes 2005) but this was not detectable after a single rotation in this study. Although the N concentrations of the crop residues were higher for the preceding legumes than preceding non-legumes, the total quantity of crop residue returned was greater for the non-legumes. This could also have contributed to the similar labile organic N concentrations among the preceding crops in the short-term. Previous findings suggest that changes in labile organic N fractions due to crop rotations may not be obvious in the short-term but rather in the long-term (Biederbeck et al. 1998; Campbell et al. 2001). Nonetheless, the results of this 1-yr study are similar to some long-term studies on the effect of crop rotation on MBN (Griffin and Porter 2004; Franchini et al. 2007; Sainju et al. 2007), POMN and LFOMN (Sharifi et al. 2008b; Sequeira and Alley 2011). In a 2-yr study in Alberta, Canada, Soon et al. (2001) found similar MBN concentrations following spring wheat, red clover (Trifolium pratense L.) green manure and field pea. Even after 20 yr, Balota et al. (2003) found similarities in MBN concentrations at 0- to 5-cm soil depth among soybean [*Glycine max* L.]–wheat, corn-wheat and cotton [Gossypium spp.]-wheat rotations in a no-till soil in Brazil. Conversely, Biederbeck et al. (2005) found greater (>60%) MBN concentrations after 6 yr at the 0- to 10-cm depth in legume green manure-wheat rotations compared with conventional fallow-wheat and continuous wheat rotations in a conventionally tilled semi-arid Canadian prairie soil. The legumes used by Biederbeck et al. (2005) were black lentil (Lens culinaris Medikus), Tangier flatpea (Lathyrus tingitanus L.), chickling vetch (Lathyrus sativus L.) and field pea (Pisum sativum L.). Furthermore, Griffin and Porter (2004) reported that the inclusion of red clover as cover crop in 2-yr potato (Solanum tuberosum L.) rotations increased the proportion of total soil N as POMN by 13–20% compared with rotations that did not contain a legume cover crop. The proportion of total soil N attributed to the labile organic N fractions was within the reported range for MBN (Campbell et al. 1991a; Willson et al. 2001; Biederbeck et al. 2005), WEON (Zsolnay 1996; Chantigny et al. 2010), POMN and LFOMN (Janzen et al. 1992; Gregorich et al. 2006; Sharifi et al. 2007b, 2008a; Sequeira and Alley 2011).

Soil mineral N concentrations varied at only two (Indian Head and Scott) of the five sites and were greater following faba bean green manure at these sites. This was attributed to the higher N<sub>2</sub>-fixation potential of faba bean (Lupwayi and Kennedy 2007) and greater quantities of crop residue N returned by faba been green manure (Unkovich and Pate 2000) resulting in greater residual soil N at the two sites. The slow decomposition of N-poor residues (non-legumes) and possible volatilization of N from the N-rich residues (legumes) on the soil surface (Schoenau and Campbell 1996) could also account for the similarities in mineral N concentrations. Another possible explanation could be due to the duration of the experiment since studies (Zebarth et al. 2009a,b) show that management of preceding crops may have limited impact on soil mineral N in the subsequent year. Finally, the sampling depth (0-15 cm) used in this study was possibly not adequate to fully account for residual N from the preceding legumes in these prairie soils (Soper and Huang 1963; Soper et al. 1971). Additional N might have been found if soil samples were taken to a depth of 60 cm, since NO<sub>3</sub>-N was probably leached below the 15-cm depth (Malhi et al. 2009).

#### **Mineralizable N Pools**

In this study, the fixed-k approach was used to estimate  $N_0$ . Wang et al. (2003) suggested using a fixed k value to eliminate the confounding effect of k on  $N_0$  thereby allowing  $N_0$  to be a single indicator of the size of the mineralizable N pool. The  $N_0$  differed among the preceding crops only at Brandon, implying that  $N_0$ was more influenced by site-specific conditions than the preceding crops (Wang et al. 2003; Dessureault-Rompre et al. 2010) since this was the first year of the study. Dessureault-Rompre et al. (2010) argued that  $N_0$  is a reflection of mineralizable N that is a function of soil properties, climatic zones and organic amendments and less responsive to recent cropping. In a 30-yr crop rotation study in a thin Black Chernozem soil,  $N_0$  was greater in legume green manure-wheat rotations than fallow-wheat-wheat rotations (Campbell et al. 1991b). Pool I, which is very labile and more responsive to recent cropping, did not vary among the preceding crops. In soils with low SON content under humid conditions in eastern Canada, Zebarth et al. (2009a) found no effect of fertilizer N management on Italian ryegrass (Lolium multiflorum Lam.) crop on labile mineralizable N, while Sharifi et al. (2009) reported higher concentrations of Pool I in 2-yr potato rotations that included legumes in comparison with non-legumes. In both of these studies, the preceding crop residues were incorporated, but were left on the soil surface in the current study. The  $N_0$ , Pool I and Pool II values were similar

to those reported in other studies using annual crops (Sharifi et al. 2007a, 2009; Nyiraneza et al. 2012), but slightly lower than in others (Campbell et al. 1991b; Sharifi et al. 2008a, 2011) where long-term crop rotations, tillage, and organic amendment history were investigated.

## Prediction of Soil N Supply

A major objective of this study was to identify parameter(s) that could be used as an indicator of soil N supply for canola production. The significant relationship between canola grain yield and N uptake suggests that N was a limiting factor across these sites. Hence, the use of grain yield as a measure of soil N supply was valid. We found that POMN was the best single predictor of soil N supply to canola in this study. However, the relationship was driven to a large extent by two clusters of data points, one cluster for the Lacombe site and the other cluster for the four remaining sites. This was due to the greater total soil N concentration at the Lacombe site relative to the other sites. Furthermore, the relationship within each site was not particularly robust. This suggests that the differences in soil N supply at each site after one rotation cycle was not accurately accounted for by POMN, most likely due to the differences in quantity of returned residues and their mineralization rates. Studies show that POMN may vary monthly or seasonally, depending on site characteristics, timing, quantity and quality of organic residue inputs and the time of soil sampling for POM extraction (Boone 1994). Hence, POMN is a dynamic pool that changes over time.

The POMN is an intermediate pool between fresh plant residues and stabilized organic N that is mostly associated with sand-sized particles in soils (Gregorich et al. 2006). The fact that POMN was the largest labile organic N fraction measured indicates that there was possibly an accumulation of POMN due to slow decomposition of crop residues resulting from the no-till practices and the sub-humid to semi-arid conditions at the sites (Biederbeck et al. 1994; Liang et al. 2004; Malhi et al. 2008, 2011b). Slow decomposition of crop residues on the soil surface in no-till systems is mainly due to enhanced aggregate formation that can protect the residues from microbial access (Six et al. 1999) and to reduced contact of crop residues with the soil particles, resulting in lower nutrient availability to microbes colonizing the surface residues (Schoenau and Campbell 1996). Additionally, reduced moisture availability in semiarid conditions can constrain decomposition. During the growing season, the accumulated POMN could have mineralized, thereby increasing soil mineral N availability, particularly in the faba bean green manure plots. Studies reported that POM is a readily available substrate for microbes and a major source of N to crops (Whalen et al. 2000; Gregorich et al. 2006). The relationship between POMN and soil N supply was better than reported in some studies (Sharifi et al. 2007a,

2008b), but weaker than in others (Spargo et al. 2011). However, these studies were conducted under wetter conditions, with different crops and over a longer period of time in comparison with the current study. This was the first study to relate POMN to soil N supply in canola production. Pre-plant soil mineral N and total soil N were also good indicators of soil N supply in this study. Other studies reported comparable results for soil mineral N as a predictor of soil N supply (Hong et al. 1990; Sharifi et al. 2007a, 2008b), while the predictions were better in others (Soper et al. 1971; Malhi et al. 1992; Nyiraneza et al. 2009; Sharifi et al. 2009). It is well documented that NO<sub>3</sub>-N measured up to the 60-cm depth plays a major role in determining soil N supply in prairie soils (Soper and Huang 1963). Soper et al. (1971) found a better relationship between barley N uptake and NO<sub>3</sub>-N measured at 61 cm ( $R^2 = 0.84$ ) than NO<sub>3</sub>-N measured at 30 cm ( $R^2 = 0.64$ ) or at 15 cm ( $R^2 = 0.32$ ). In spite of these findings, the 0- to 15-cm sampling depth was used in our study since soil biochemical activity is more pronounced at the soil surface. Hence, most indices of N availability reflect surface processes. In addition, the potential to detect differences in soil mineral N concentration and other parameters, such as POMN, in these no-till sites would be greatly diluted if greater sampling depths were used, particularly as the crop residues were not incorporated in the soil. Nevertheless, regardless of the sampling depth, the use of soil mineral N or NO<sub>3</sub>-N to predict soil N supply could be problematic since soil mineral N concentration changes rapidly, making the values sensitive to sampling date (Sharifi et al. 2007a).

 $N_0$  was not an accurate predictor of soil N supply in this study, suggesting that  $N_0$  may not be a good estimator of the fertilizer N needs of crops. Other studies reported similar findings and suggested that  $N_0$  cannot accurately simulate microbial activity, or reflect crop effects and possible N losses under field conditions (Sharifi et al. 2007a; Nyiraneza et al. 2012). In addition, Campbell et al. (1991b) argued that the initial potential rate of N mineralization  $(N_0k)$  was a more sensitive parameter than  $N_0$  and may be a more useful index for identifying the beneficial effects of legume green manures in cereal rotations. Conversely, Spargo et al. (2011) reported that  $N_0$  was a reliable predictor of corn grain yield (r = 0.87, P < 0.001) and N uptake (r = 0.83, P < 0.001) across different organic and conventional cropping systems at one site in Maryland, USA.

Combining soil physio-chemical properties and climatic variables (e.g., precipitation) gave the best prediction of soil N supply to canola. Up to 68 and 71% of the variations in yield and N uptake, respectively, were explained when PC<sub>2</sub>, PC<sub>3</sub> and PC<sub>4</sub> were added to the statistical model. Taking into account the loading values, PC<sub>2</sub> was interpreted as POMN across a temperature gradient. The PC<sub>3</sub> represented soil texture effect across a precipitation gradient, while PC<sub>4</sub> represented plantavailable N. The PC<sub>1</sub> represented a minor source of mineralizable N in which soil pH played a significant role. Because  $PC_1$  accounted for only 7% of the total variability in grain yield, it can be concluded that POMN and soil mineral N accounted for much of the variation in soil N supply to canola across these no-till sites with soil texture influencing the magnitude of N supply.

## CONCLUSION

Although some effects of preceding legume crops on soil N supply were identified in this study, the magnitude of the impact varied across legume species, soil properties and climatic conditions. This study demonstrated that soil MBN, WEON, POMN and LFOMN may not be responsive to preceding crops in the short-term in no-till systems. Due to possible accumulation and subsequent mineralization of POMN, the results indicate that POMN and mineral N are relatively good predictors of soil N supply to canola under sub-humid and semiarid conditions in western Canada and that soil texture also influences the soil N supply. This suggests that canola N fertilizer recommendations may be improved by taking into account POMN and pre-plant soil mineral N concentrations as well as soil texture. The POMN was the largest pool of labile organic N in soils under no-tillage and may represent a fraction of SON that is decomposed early in the growing season, thereby furnishing plant-available N to canola during early vegetative growth and when N requirements are greatest (five-leaf stage to full bloom; Canola Council of Canada 2012b). More research on the mechanisms controlling the formation and dynamics of POMN are needed to better understand its role in agricultural soils for improved N management.

## ACKNOWLEDGEMENTS

Funding was provided by the Sustainable Agricultural Environment Systems (SAGES) program of Agriculture and Agri-Food Canada. The first author was also supported by a grant from Centre Sève. Technical assistance was provided by Sylvie Côté, Claude Lévesque, Ginette Decker, Aleksandra Spasojevic, Randy Shiplack and Kevin Willoughby.

Balota, E. L., Colozzi-Filho, A., Andrade, D. S. and Dick, R. P. 2003. Microbial biomass in soils under different tillage and crop rotation systems. Biol. Fertil. Soils 38: 15–20.

**Biederbeck, V. O., Janzen, H. H., Campbell, C. A. and Zentner, R. P. 1994.** Labile soil organic matter as influenced by cropping practices in an arid environment. Soil Biol. Biochem. **26**: 1647–1656.

Biederbeck, V. O., Rice, W. A., van Kessel, C., Bailey, L. D. and Huffman, E. C. 1996. Present and potential future nitrogen gains from legumes in major soil zones of the prairies. Proc. Soils and Crops 1996, University of Saskatchewan, SK.

**Biederbeck, V. O., Campbell, C. A., Rasiah, V., Zentner, R. P. and Wen, G. 1998.** Soil quality attributes as influenced by annual legumes used as green manure. Soil Biol. Biochem. **30**: 1177–1185.

**Biederbeck, V. O., Zentner, R. P. and Campbell, C. A. 2005.** Soil microbial populations and activities as influenced by legume green fallow in a semiarid climate. Soil Biol. Biochem. **37**: 1775–1784.

Boone, R. D. 1994. Light-fraction soil organic matter: origin and contribution to net nitrogen mineralization. Soil Biol. Biochem. 26: 1459–1468.

**Brookes, P. C. 2001.** The soil microbial biomass: concept, measurement and applications in soil ecosystem research. Microbes Environ. **16**: 131–140.

**Brookes, P. C., Landman, A., Pruden, G. and Jenkinson, D. S. 1985.** Chloroform fumigation and the release of soil nitrogen: A rapid direct extraction method to measure microbial biomass nitrogen in soil. Soil Biol. Biochem. **17**: 837–842.

Cabrera, M. L. and Beare, M. H. 1993. Alkaline persulfate oxidation for determining total nitrogen in microbial biomass extracts. Soil Sci. Soc. Am. J. 57: 1007–1012.

Campbell, C. A., Biederbeck, V. O., Zentner, R. P. and Lafond, G. P. 1991a. Effect of crop rotations and cultural practices on soil organic matter, microbial biomass and respiration in a thin black Chernozem. Can. J. Soil Sci. 71: 363–376.

Campbell, C. A., Lafond, G. P., Leyshon, A. J., Zentner, R. P. and Janzen, H. H. 1991b. Effect of cropping practices on the initial potential rate of N mineralization in a thin Black Chernozem. Can. J. Soil Sci. 71: 43–53.

Campbell, C. A., Lafond, G. P., van den Bygaart, A. J., Zentner, R. P., Lemke, R., May, W. E. and Holzapfel, C. B. 2011. Effect of crop rotation, fertilizer and tillage management on spring wheat grain yield and N and P content in a thin Black Chernozem: A long-term study. Can. J. Plant Sci. 91: 467–483.

Campbell, C. A., Selles, F., Lafond, G. P., Biederbeck, V. O. and Zentner, R. P. 2001. Tillage–fertilizer changes: Effect on some soil quality attributes under long-term crop rotations in a thin Black Chernozem. Can. J. Soil Sci. 81: 157–165.

Canola Council of Canada. 2012a. Provincial acreages and yields. [Online] Available: http://www.canolacouncil.org/markets-stats/statistics/harvest-acreage/ [2012 Oct. 06].

**Canola Council of Canada. 2012b.** Good time for top dressing. [Online] Available: http://www.canolawatch.org/2012/05/31/ good-time-for-top-dressing/ [2012 Oct. 22].

Carson, J. A., Harapiak, J. T., Hennig, A. M. F., Janke, W. E., Nyborg, M., Penney, D. C. and Walker, D. R. 1974. Use of soil testing to predict nitrogen fertilizer needs of barley and rapeseed in Alberta. Proc. Proc. Soil fertility workshop, Saskatoon, SK. Feb. 1974.

Chantigny, M. H. 2003. Dissolved and water-extractable organic matter in soils: A review on the influence of land use and management practices. Geoderma 113: 357–380.

Chantigny, M. H., Angers, D. A., Kaiser, K. and Kalbitz, K. 2008. Extraction and characterization of dissolved organic matter. Pages 617–635 *in* M. R. Carter and E. G. Gregorich, eds. Soil sampling and methods of analysis. 2nd ed. Canadian Society of Soil Science, CRC Press, Boca Raton, FL.

Chantigny, M. H., Curtin, D., Beare, M. H. and Greenfield, L. G. 2010. Influence of temperature on water-extractable organic matter and ammonium production in mineral soils. Soil Sci. Soc. Am. J. 74: 517–524.

**Curtin, D. and Campbell, C. A. 2008.** Mineralizable nitrogen. Pages 599–606 *in* M. R. Carter and E. G. Gregorich, eds. Soil sampling and methods of analysis. 2nd ed. CRC Press, Boca Raton, FL. Cutforth, H., McConkey, B., Brandt, S., Gan, Y., Lafond, G., Angadi, S. and Judiesch, D. 2009. Fertilizer N response and canola yield in the semiarid Canadian prairies. Can. J. Plant Sci. 89: 501–503.

Dessureault-Rompre, J., Zebarth, B. J., Burton, D. L., Sharifi, M., Cooper, J. M. and Grant, C. A. 2010. Relationships among mineralizable soil nitrogen, soil properties, and climatic indices. Soil Sci. Soc. Am. J. 74: 1218–1227.

**Dosdall, L. M., Harker, K. N., O'Donovan, J. T., Blackshaw, R. E., Kutcher, H. R., Gan, Y. and Johnson, E. N. 2012.** Crop sequence effects on root maggot (Diptera: Anthomyiidae: *Delia* spp.) infestations in canola. J. Econ. Entomol. **105**: 1261–1267.

Fox, R. H. and Piekielek, W. P. 1978. Field testing of several nitrogen availability indexes. Soil Sci. Soc. Am. J. 42: 747–750.

Franchini, J. C., Crispino, C. C., Souza, R. A., Torres, E. and Hungria, M. 2007. Microbiological parameters as indicators of soil quality under various soil management and crop rotation systems in southern Brazil. Soil Tillage Res. 92: 18–29.

**Gregorich, E. G. and Beare, M. H. 2008.** Physically uncomplexed organic matter. Pages 607–616 *in* M. R. Carter and E. G. Gregorich, eds. Soil sampling and methods of analysis. 2nd ed. Canadian Society of Soil Science, CRC Press, Boca Raton, FL.

Gregorich, E. G., Beare, M. H., McKim, U. F. and Skjemstad, J. O. 2006. Chemical and biological characteristics of physically uncomplexed organic matter. Soil Sci. Soc. Am. J. 70: 975–985.

Gregorich, E. G., Monreal, C. M., Schnitzer, M. and Schulten, H. R. 1996. Transformation of plant residues into soil organic matter: chemical characterization of plant tissue, isolated soil fractions, and whole soils. Soil Sci. 161: 680–693.

Griffin, T. S. and Porter, G. A. 2004. Altering soil carbon and nitrogen stocks in intensively tilled two-year rotations. Biol. Fertil. Soils **39**: 366–374.

Harker, K. N., O'Donovan, J. T., Turkington, T. K., Blackshaw, R. E., Lupwayi, N. Z., Smith, E. G., Klein-Gebbinck, H., Dosdall, L. M., Hall, L. M., Willenborg, C. J. and others 2012. High-yield no-till canola production on the Canadian prairies. Can. J. Plant Sci. 92: 221–233.

Haynes, R. J. 2005. Labile organic matter fractions as central components of the quality of agricultural soils: An overview. Adv. Agron. 85: 221–268.

Hendershot, W. H., Lalande, H. and Duquette, M. 2008. Soil reaction and exchangeable acidity. Pages 173–178 *in* M. R. Carter and E. G. Gregorich, eds. Soil sampling and methods of analysis, 2nd ed. Canadian Society of Soil Science, CRC Press, Boca Raton, FL.

Hong, S., Fox, R. and Piekielek, W. 1990. Field evaluation of several chemical indexes of soil nitrogen availability. Plant Soil 123: 83–88.

Janzen, H. H., Campbell, C. A., Brandt, S. A., Lafond, G. P. and Townley-Smith, L. 1992. Light-fraction organic matter in soils from long-term crop rotations. Soil Sci. Soc. Am. J. 56: 1799–1806.

Kaiser, H. 1974. An index of factorial simplicity. Psychometrika 39: 31–36.

**Kroetsch, D. and Wang, C. 2008.** Particle size distribution. Pages 713–725 *in* M. R. Carter and E. G. Gregorich, eds. Soil sampling and methods of analysis. 2nd ed. CRC Press, Boca Raton, FL. Kumar, K. and Goh, K. M. 1999. Crop residues and management practices: Effects on soil quality, soil nitrogen dynamics, crop yield, and nitrogen recovery. Adv. Agron. 68: 197–319.

Liang, B. C., McConkey, B. G., Campbell, C. A., Curtin, D., Lafond, G. P., Brandt, S. A. and Moulin, A. P. 2004. Total and labile soil organic nitrogen as influenced by crop rotations and tillage in Canadian prairie soils. Biol. Fertil. Soils 39: 249–257. Lupwayi, N. Z. and Kennedy, A. C. 2007. Grain legumes in Northern Great plains: Impacts on selected biological soil processes. Agron. J. 99: 1700–1709.

Malhi, S. S. and Gill, K. S. 2004. Placement, rate and source of N, seedrow opener and seeding depth effects on canola production. Can. J. Plant Sci. 84: 719–729.

Malhi, S. S., Brandt, S. A., Kutcher, H. R. and Ulrich, D. 2011a. Effects of broad-leaf crop frequency and fungicide application in various rotations on nitrate nitrogen and extractable phosphorus in a dark brown soil. Commun. Soil Sci. Plant Anal. 42: 2795–2812.

Malhi, S. S., Brandt, S., Lemke, R., Moulin, A. and Zentner, R. 2009. Effects of input level and crop diversity on soil nitrate-N, extractable P, aggregation, organic C and N, and nutrient balance in the Canadian Prairie. Nutr. Cycling Agroecosyst. 84: 1–22.

Malhi, S. S., Moulin, A. P., Johnston, A. M. and Kutcher, H. R. 2008. Short-term and long-term effects of tillage and crop rotation on soil physical properties, organic C and N in a Black Chernozem in northeastern Saskatchewan. Can. J. Soil Sci. 88: 273–282.

Malhi, S. S., Nyborg, M. and Heaney, D. J. 1992. Potential for nitrogen mineralization in central Alberta soils. Nutr. Cycling Agroecosyst. 32: 321–325.

Malhi, S. S., Nyborg, M., Goddard, T. and Puurveen, D. 2011b. Long-term tillage, straw management and N fertilization effects on quantity and quality of organic C and N in a Black Chernozem soil. Nutr. Cycling Agroecosyst. 90: 227–241.

Nyiraneza, J., N'Dayegamiye, A., Chantigny, M. H. and Laverdière, M. R. 2009. Variations in corn yield and nitrogen uptake in relation to soil attributes and nitrogen availability indices. Soil Sci. Soc. Am. J. 73: 317–327.

Nyiraneza, J., Zebarth, B. J., Ziadi, N., Sharifi, M., Burton, D. L., Drury, C. F., Bittman, S. and Grant, C. A. 2012. Prediction of soil nitrogen supply in corn production using soil chemical and biological indices. Soil Sci. Soc. Am. J. 76: 925–935.

Sainju, U. M., Lenssen, A., Caesar-Thonthat, T. and Waddell, J. 2007. Dryland plant biomass and soil carbon and nitrogen fractions on transient land as influenced by tillage and crop rotation. Soil Tillage Res. 93: 452–461.

**SAS Institute, Inc. 2004.** SAS OnlineDoc. Version 9.1.3. SAS Institute, Inc., Cary, NC.

Schoenau, J. J. and Campbell, C. A. 1996. Impact of crop residues on nutrient availability in conservation tillage systems. Can. J. Plant Sci. 76: 621–626.

Sequeira, C. H. and Alley, M. M. 2011. Soil organic matter fractions as indices of soil quality changes. Soil Sci. Soc. Am. J. 75: 1766–1773.

Sharifi, M., Zebarth, B., Burton, D., Grant, C., Porter, G., Cooper, J., Leclerc, Y., Moreau, G. and Arsenault, W. 2007a. Evaluation of laboratory-based measures of soil mineral nitrogen and potentially mineralizable nitrogen as predictors of field-based indices of soil nitrogen supply in potato production. Plant Soil 301: 203–214. Sharifi, M., Zebarth, B. J., Burton, D. L., Grant, C. A. and Cooper, J. M. 2007b. Evaluation of some indices of potentially mineralizable nitrogen in soil. Soil Sci. Soc. Am. J. 71: 1233–1239.

Sharifi, M., Zebarth, B. J., Burton, D. L., Grant, C. A., Bittman, S., Drury, C. F., McConkey, B. G. and Ziadi, N. 2008a. Response of potentially mineralizable soil nitrogen and indices of nitrogen availability to tillage system. Soil Sci. Soc. Am. J. 72: 1124–1131.

Sharifi, M., Zebarth, B. J., Burton, D. L., Grant, C. A. and Porter, G. A. 2008b. Organic amendment history and crop rotation effects on soil nitrogen mineralization potential and soil nitrogen supply in a potato cropping system. Agron. J. 100: 1562–1572.

Sharifi, M., Zebarth, B. J., Burton, D. L., Rodd, V. and Grant, C. A. 2011. Long-term effects of semisolid beef manure application to forage grass on soil mineralizable nitrogen. Soil Sci. Soc. Am. J. 75: 649–658.

Sharifi, M., Zebarth, B. J., Porter, G. A., Burton, D. L. and Grant, C. A. 2009. Soil mineralizable nitrogen and soil nitrogen supply under two-year potato rotations. Plant Soil 320: 267–279.

Six, J., Elliott, E. T. and Paustian, K. 1999. Aggregate and soil organic matter dynamics under conventional and no-tillage systems. Soil Sci. Soc. Am. J. 63: 1350–1358.

Skjemstad, J. and Baldock, J. 2008. Total and organic carbon. Pages 225–237 *in* M. R. Carter and E. G. Gregorich, eds. Soil sampling and methods of analysis. 2nd ed. Canadian Society of Soil Science, CRC Press, Boca Raton, FL.

Soon, Y. K. and Arshad, M. A. 2004. Tillage, crop residue and crop sequence effects on nitrogen availability in a legume-based cropping system. Can. J. Soil Sci. 84: 421–430.

Soon, Y. K., Clayton, G. W. and Rice, W. A. 2001. Tillage and previous crop effects on dynamics of nitrogen in a wheat-soil system. Agron. J. **93**: 842–849.

Soper, R. J. and Huang, P. M. 1963. The effect of nitrate nitrogen in the soil profile on the response of barely to fertilizer nitrogen. Can. J. Soil Sci. 43: 359–358.

Soper, R. J., RacZ, G. J. and Fehr, P. I. 1971. Nitrate nitrogen in the soil as a means of predicting the fertilizer nitrogen requirements of barley. Can. J. Soil Sci. 51: 45–49.

Spargo, J. T., Cavigelli, M. A., Mirsky, S. B., Maul, J. E. and Meisinger, J. J. 2011. Mineralizable soil nitrogen and labile soil organic matter in diverse long-term cropping systems. Nutr. Cycling Agroecosyst. 90: 253–266.

St. Luce, M., Whalen, J. K., Ziadi, N. and Zebarth, B. J. 2011. Nitrogen dynamics and indices to predict soil nitrogen supply in humid temperate soils. Adv. Agron. 112: 55–102.

Stanford, G. and Smith, S. J. 1972. Nitrogen mineralization potentials of soils. Soil Sci. Soc. Am. J. 36: 465–472.

Thorup-Kristensen, K., Magid, J. and Jensen, L. S. 2003. Catch crops and green manures as biological tools in nitrogen management in temperate zones. Adv. Agron. 79: 227–302.

**Unkovich, M. J. and Pate, J. S. 2000.** An appraisal of recent field measurements of symbiotic N2 fixation by annual legumes. Field Crop Res. **65**: 211–228.

Voroney, R. P., Brookes, P. C. and Beyaert, R. P. 2008. Soil microbial biomass C, N, P, and S. Pages 637–651 *in* M. R. Carter and E. G. Gregorich, eds. Soil sampling and methods of soil analysis. 2nd ed. Canadian Society of Soil Science, CRC Press, Boca Raton, FL.

Walley, F., Yates, T., van Groenigen, J.-W. and van Kessel, C. 2002. Relationships between soil nitrogen availability indices, yield, and nitrogen accumulation of wheat. Soil Sci. Soc. Am. J. 66: 1549–1561.

Wang, W. J., Smith, C. J. and Chen, D. 2003. Towards a standardised procedure for determining the potentially mineralisable nitrogen of soil. Biol. Fertil. Soils **37**: 362–374.

Whalen, J. K., Bottomley, P. J. and Myrold, D. D. 2000. Carbon and nitrogen mineralization from light- and heavyfraction additions to soil. Soil Biol. Biochem. 32: 1345–1352.

Willson, T. C., Paul, E. A. and Harwood, R. R. 2001. Biologically active soil organic matter fractions in sustainable cropping systems. Appl. Soil Ecol. 16: 63–76.

Wuensch, K. L. 2004. Principle components analysis. [Online] Available: http://core.ecu.edu/psyc/wuenschk/MV/FA/PCA. doc [2012 Oct. 03].

Zebarth, B. J. and Milburn, P. H. 2003. Spatial and temporal distribution of soil inorganic nitrogen concentration in potato hills. Can. J. Soil Sci. 83: 183–195.

Zebarth, B. J., Arsenault, W. J., Moorehead, S., Kunelius, H. T. and Sharifi, M. 2009a. Italian ryegrass management effects on nitrogen supply to a subsequent potato crop. Agron. J. 101: 1573–1580.

Zebarth, B. J., Scott, P. and Sharifi, M. 2009b. Effect of straw and fertilizer nitrogen management for spring barley on soil nitrogen supply to a subsequent potato crop. Am. J. Potato Res. 86: 209–217.

Zentner, R. P., Campbell, C. A., Biederbeck, V. O., Miller, P. R., Selles, F. and Fernandez, M. R. 2001. In search of a sustainable cropping system for the semiarid Canadian prairies. J. Sustain. Agric. 18: 117–136.

Zotarelli, L., Zatorre, N. P., Boddey, R. M., Urquiaga, S., Jantalia, C. P., Franchini, J. C. and Alves, B. J. R. 2012. Influence of no-tillage and frequency of a green manure legume in crop rotations for balancing N outputs and preserving soil organic C stocks. Field Crop Res. 132: 185–195. Zsolnay, Á. 1996. Dissolved humus in soil waters. Pages 171–223 *in* A. Piccolo, ed. Humic substances in terrestrial ecosystems. Elsevier, Amsterdam, the Netherlands.